

N-Acetylcysteine: A New Approach to Anti-HIV Therapy

MARIO ROEDERER, STEPHEN W. ELA, FRANK J.T. STAAL, LEONORE A. HERZENBERG, and
 LEONARD A. HERZENBERG

ABSTRACT

Several investigators have implicated depletion of glutathione (GSH) and production of reactive oxygen intermediates (ROIs) in the regulation of the human immunodeficiency virus (HIV). We have shown directly that *N*-acetylcysteine (NAC) blocks HIV expression in chronic and acute infection models, and HIV replication in normal peripheral blood mononuclear cells. NAC is a cysteine prodrug which maintains intracellular thiol levels during oxidative stress and replenishes depleted GSH. The observed antiviral effect of NAC is due to inhibition of viral stimulation by ROIs, which are produced in response to inflammatory cytokines.

We have also shown that HIV-infected individuals have decreased intracellular GSH levels in their circulating T cells. Since GSH is the major protection against the production of ROIs, we hypothesize that the observed decrease is due to a chronic oxidative stress induced by continual exposure to elevated levels of inflammatory cytokines. Together, these results provide a rationale for clinical trials testing the efficacy of GSH-replenishing drugs such as NAC in the treatment of AIDS. NAC is different than many other antiviral drugs in that it inhibits host-mediated stimulation of viral replication arising in normal immune responses, and may thereby extend latency. In addition, it inhibits the action of inflammatory cytokines which may mediate cachexia, thereby raising the possibility that it may alleviate the deleterious wasting that accompanies late stage AIDS.

INTRODUCTION

DROGE AND COLLEAGUES FIRST SHOWED that human immunodeficiency virus-positive (HIV⁺) individuals have decreased levels of acid-soluble thiols, in particular cysteine and glutathione (GSH), in their plasma and leukocytes.¹ This observation suggests that oxidative stress may play an important role in the progression of acquired immunodeficiency syndrome (AIDS): because GSH is the major intracellular defense against the production of reactive oxygen intermediates (ROIs), overproduction of such oxidants depletes GSH. In addition, they showed that glutamate levels were increased in plasma. Glutamate inhibits the cellular uptake of cystine, used in GSH synthesis. Crystal and colleagues confirmed the depletion of plasma GSH, and further showed that there is a depression of the GSH levels in the bronchoalveolar lavage fluid of HIV⁺ individuals.² We have shown that the decrease in intracellular GSH in leukocytes is confined primarily to the T-cell subsets.³⁻⁶ Finally, Smith et al. have shown that vertical transmission of HIV to children also results in decreased GSH levels.⁷

We suggested that the decline in GSH in HIV⁺ individuals might be due to chronic exposure to inflammatory cytokines.⁴ Fauci and colleagues suggested that inflammatory cytokines such as tumor necrosis factor α (TNF), interleukin (IL)-1, IL-6, and granulocyte-macrophage colony stimulating factor (GM-CSF) play central roles in the progression of HIV.⁸⁻¹⁰ Several laboratories have shown that HIV⁺ individuals have elevated production and levels of inflammatory cytokines.¹¹⁻¹⁷ Besides directly stimulating transcription and replication of HIV, these cytokines cause an oxidative stress at both a systemic and cellular level. Thus, chronic exposure to elevated levels of cytokines may result in depletion of the major antioxidant defense, intracellular GSH.

These observations led to the suggestion that treatment of the HIV infection (and other syndromes in which inflammatory cytokines are overproduced) should include agents to restore systemic and intracellular GSH.¹⁸⁻²¹ Over the past two years, several studies showed that GSH-replenishing drugs, such as *N*-acetylcysteine (NAC), glutathione ethyl ester, and GSH itself, have profound effects on the regulation of inflammatory

Department of Genetics, Stanford University, Stanford, CA 94305.

cytokines: for instance, these compounds strongly inhibit the stimulation of HIV (in both acute infections and chronic infections) by TNF, IL-1, IL-6, (GM-CSF), and phorbol ester.^{18-20,22,23}

In this report, we review the basic observations which have led to the hypothesis that oxidative stress plays a critical role in AIDS, and that drugs which are GSH precursors can alleviate this stress. On the basis of these results, we strongly recommend that treatment of AIDS include therapies aimed at restoring depleted GSH levels.

INFLAMMATORY CYTOKINES DIRECTLY STIMULATE HIV

HIV has taken advantage of signaling pathways by inflammatory cytokines to regulate its own transcription and replication. The HIV-1 long terminal repeat (LTR), the promoter and enhancer region for viral transcription, has two binding sites for the enhancer protein NF- κ B,²⁴ and the HIV-2 LTR has one such site. NF- κ B is a cytoplasmic factor which, upon cellular stimulation, translocates to the nucleus in an active form.^{25,26} NF- κ B activation is sufficient for stimulation of the HIV LTR by TNF and phorbol myristate acetate (PMA).^{24,27-30} Once active and in the nucleus, it can increase transcription of the viral genome tenfold.

Through the stimulation of transcription of HIV, TNF can activate otherwise latent virus.^{8,31} Fauci and colleagues have developed two chronically infected cell lines, ACH-2 (T cell) and UI (promonocyte), which serve as models for latency.^{8,27} These cell lines do not produce virions unless stimulated; agents which can induce virus in these lines include TNF, PMA, GM-CSF, and IL-6.⁹

Many cytokines will synergize with each other to give significantly greater induction than occurs with only one. This synergism occurs at the level of induction of NF- κ B,^{23,32} mRNA and protein^{18,22,32} production of virus in chronically infected cell lines,⁹ and replication of virus in acutely infected PBMC.^{18,22} Synergism is extremely important from the standpoint that low production of several cytokines (at levels undetectable in blood sera) may result in as great a stimulation of HIV as high production of a single cytokine.

INFLAMMATORY CYTOKINES CAUSE OXIDATIVE STRESS THAT CAN BE COUNTERED BY INTRACELLULAR GSH

Production of ROIs is a normal outcome of stimulation of many types of cells. The most dramatic is that by neutrophils after stimulation by, for example, TNF.³³⁻³⁵ This "respiratory burst" includes the production of a variety of oxygen radicals, such as superoxide anion and hydroxyl radical. However, other cell types also produce ROIs in response to stimulation (reviewed by Cross and Jones³⁶), including B lymphocytes,^{37,38} T lymphocytes,^{39,40} endothelial cells,⁴¹ and fibroblasts.^{42,43} PMA also causes the production of ROIs.^{33,44} While TNF by itself can induce oxidant production, it is far more potent in combination with certain other agents (synergistic stimulation). In combination with interferon- γ (IFN γ) or IFN α , a significantly higher production of ROIs is possible.⁴⁵

The main defense against the potentially damaging ROIs is intracellular GSH. This cysteine-containing tripeptide is found in virtually all eukaryotic organisms in millimolar concentrations in the cytoplasm and mitochondria. It is the largest source of free thiol inside cells, and thereby regulates the redox potential within the cell. The enzyme-catalyzed reduction (neutralization) of ROIs consumes GSH (which is converted to GSSG). Subsequently, GSSG is either exported from the cell or enzymatically reduced to GSH by glutathione reductase.^{46,47}

Stimulation of polymorphonuclear cells with PMA, inducing a respiratory burst, can consume as much as 50% of the intracellular GSH.⁴⁸ Bilzer and Lauterburg showed that this was also true for stimulation with TNF; furthermore, they showed that the continued resynthesis of GSH is critical in maintaining the intracellular GSH levels after stimulation.⁴⁹

Stimulation of cells may overcome the protective effects of intracellular GSH.^{48,50} However, addition of an exogenous thiol source can restore the intracellular reducing capability. Zimmerman et al. have shown that NAC, which can restore intracellular GSH and directly scavenge ROIs,^{51,52} neutralizes the toxic effects of TNF both on tumor cell lines and in vivo, in rats injected with lethal doses of TNF.^{50,53} By virtue of its ability to replace GSH, NAC is the antidote to acetaminophen overdose in humans,⁵⁴⁻⁵⁶ a condition which leads to fatal depletion of GSH in the liver.

INTRACELLULAR THIOL LEVELS REGULATE LYMPHOCYTE FUNCTION

While the observations summarized above suggest that intracellular GSH levels directly modulate stimulation by TNF (and other agents), it is difficult to separate the effects of increased ROIs from decreased GSH (and vice versa). Since production of ROIs leads to consumption of GSH, and a depletion of GSH could lead to increased levels of intracellular ROIs, assignment of specific roles for one or the other must be made with caution. Nevertheless, a considerable amount is known about the roles that these reducing and oxidizing species play in cellular functions.

Feedback mechanisms maintain intracellular GSH levels.⁵⁷ Thus, the most effective way to lower GSH levels is to incubate cells in the presence of buthionine sulfoximine (BSO), an inhibitor of γ -glutamylcysteine synthetase.^{58,59} Incubation of dividing cells with BSO for several days can deplete as much as 90% of intracellular GSH. While maintenance of GSH levels with exogenously added NAC protects cells against TNF cytotoxicity,⁵⁰ depletion of GSH by BSO potentiates TNF cytotoxicity.⁶⁰

Wedner and colleagues first showed that depletion of GSH inhibited T-cell proliferation.⁶¹ In several studies, Fidelus et al. also showed the critical need for adequate intracellular GSH levels for T-cell proliferation.⁶²⁻⁶⁴ Reductions of GSH by 10-40% in T cells completely inhibited T-cell activation.⁶⁵ Messina and Lawrence showed that a 40% decrease in intracellular GSH inhibited cell cycle progression in PBMC. However, even a 90% decrease in intracellular GSH did not inhibit the stimulated expression of IL-2 receptor or secretion of IL-2.⁶⁶ (This is not surprising in view of the dependence of IL-2 receptor expression on activation of NF- κ B.⁶⁷⁻⁶⁹ this activation is more likely under conditions of GSH depletion—see below.)

Gmunder et al. confirmed this result,⁷⁰ and further suggested that some processes are GSH dependent, while others are cysteine dependent.⁷¹ There seemed to be no linear correlation between the degree of GSH depletion and the inhibition observed.⁶⁵

Wong et al. used a novel approach to investigate the interrelationship of ROIs and stimulation: cloning and overexpression of either manganese superoxide dismutase (MnSOD) to try to reduce ROI levels, or overexpression of the MnSOD antisense mRNA to increase ROI levels.⁷² Indeed, overexpression of the enzyme leads to inhibition of the effects of TNF, while overexpression of the antisense RNA leads to increased sensitivity to TNF (potentiation). There is also a crucial involvement of HIV in this *in vitro* system. In normal cells, TNF activates expression of MnSOD;^{72,73} however, in HIV-infected cells, this activation is suppressed.⁷⁴ Furthermore, HIV infection sensitizes cells to stresses such as heat and radiation.⁷⁴

While depletion of GSH leads to inhibition of some T-cell functions (but increased sensitivity to inflammatory cytokines), supplementation can augment other functions, both *in vitro* and *in vivo*. Exogenously added GSH augments lymphocyte proliferation in response to lectin.^{75,76} Oxothiazolidine 4-carboxylate (OTC), a cysteine precursor which increases GSH levels, acts synergistically with concanavalin A to stimulate T cells.⁶² Finally, Droge et al. showed that increasing previously lowered GSH levels in mice augments the activation of cytolytic T cells, demonstrating the importance of GSH levels *in vivo*.⁷⁷

We have shown that GSH supplementation through addition of NAC can completely inhibit inflammatory stimulations of HIV replication. NAC inhibits the stimulated expression of the HIV LTR¹⁸ through the inhibition of activation of NF- κ B.^{20,23,78} This inhibition results in blocking of viral replication in acutely infected cells^{18,22} and chronically infected cells.^{19,22} NAC not only inhibited stimulation of viral replication by TNF and PMA, but also by IL-6¹⁹ and GM-CSF (Poli and Fauci, personal communication).

Recently, Baeuerle's laboratory has confirmed and extended our observations. They have shown that a wide variety of agents which induce NF- κ B through distinct pathways (e.g., cycloheximide, double-stranded RNA, calcium ionophore, TNF, PMA, IL-1, LPS, and lectin) are all inhibited by addition of NAC.⁷⁸ This suggests that these stimulatory agents might all activate NF- κ B by the same mechanism involving a NAC-sensitive signaling step.

Perhaps most significantly, Baeuerle's group showed that hydrogen peroxide (H₂O₂) can directly and specifically activate NF- κ B in a Jurkat T-cell subclone. This activation leads to stimulation of HIV replication in latently infected Jurkat T cells.⁷⁸ H₂O₂ also stimulated viral production in latently infected U1 promonocytes.⁷⁹ These results suggest that activation of HIV *in vivo* might be accomplished not only by the stimulation of production of ROIs in infected cells, but also by production of ROIs in uninfected cells (e.g., granulocytes) which then diffuse into neighboring infected cells. While Frei et al. found an upper limit of 0.25 μ M for H₂O₂ in the blood plasma of healthy subjects,⁸⁰ it is probable that this value is higher under conditions of inflammatory stress, especially in local cellular environments.

These results confirm the observations of Fidelus, who showed that the production of oxygen radicals is not only a positive signal in T-cell activation, it is necessary for some

signaling pathways (e.g., PMA).⁶⁴ Thus, ROIs may be commonly-used second messengers for inflammatory stimulations as induced by, for example, TNF and IL-1.^{36,78} Since H₂O₂ can activate HIV in the absence of other agents, and synergizes with some stimulatory agents,⁷⁸ ROIs must be assigned an important stimulatory role in the progression of AIDS.

INFLAMMATORY AND OXIDATIVE STRESSES ACCOMPANY AIDS

In apparent contradiction to the immunosuppressed state of individuals infected with HIV are the observations that their sera have elevated levels of several cytokines. While some studies showed significantly elevated levels of serum TNF,¹¹⁻¹⁶ other studies did not show significant elevations (e.g., Fuchs et al. found no significant elevation in a retrospective study, but pointed out that instability of TNF in serum samples may lead to this discrepancy⁸¹). Voth et al. showed that TNF mRNA in the PBMC from HIV⁺ individuals has a longer half-life than that in uninfected individuals;⁸² this may account for increased TNF production by resting alveolar macrophages from these individuals.^{83,84} Other than TNF, elevated levels of IL-1 α ,¹⁶ IL-1 β ,¹⁷ IL-2,¹⁶ IL-6,^{17,85} IFN- α ,^{15,86} and IFN- γ ⁸⁷ have been measured in sera from HIV⁺ individuals. Furthermore, these individuals have hypertriglyceridemia, probably due to elevated cytokine levels.⁸⁸ These results suggest that concomitant with the HIV infection is chronic stimulation by a variety of inflammatory agents.

A strong indication that an inflammatory stress accompanies AIDS is the elevated level of plasma neopterin.^{89,90} Macrophages synthesize neopterin principally in response to IFN- γ stimulation; thus, neopterin levels serve as a marker of the status of cell-mediated immunity.⁹⁰ Not only is the level of neopterin correlated with the progression of the disease, but it is also a strong predictor for subsequent progression to AIDS.^{81,91-94}

Another indication that inflammatory stress accompanies AIDS is the chronic depletion of GSH (and other low-molecular weight thiols). HIV⁺ individuals have depleted levels of acid-soluble thiols in the plasma, and depleted levels of GSH in plasma and lung epithelial lining fluid.^{1,2} The chronic depletion of GSH is consistent with the observations of increased cytokine levels and that GSH levels (in T cells) decrease after stimulation *in vitro*.^{63,75,95}

Recently, Droge et al. have followed up on their earlier studies by showing that thiol depletion also occurs in a primate model for AIDS, that of simian immunodeficiency virus (SIV) infection of macaques.⁹⁶ This last report is extremely important from several standpoints: it suggests that the chronic thiol depletion is a general result of retroviral infections; it shows that the decrease in acid-soluble thiols can occur within a week of infection (we have seen decreased levels of GSH in a patient 3 weeks after infection);⁶ and it suggests that the macaque SIV model may be useful in testing drugs targeted at restoring GSH.

Using a flow cytometric assay for intracellular GSH simultaneously with cell identification by membrane immunofluorescence, we have quantitated the GSH levels in subsets of PBMC.^{4,6} Our results are summarized as follows: (1) Intracellular GSH levels fall within a narrow range for each PBMC subset from normal individuals;³ (2) HIV⁺ individuals have 30-40% decreased GSH levels in both CD4 and CD8 T cells;⁶

(3) this decrease is due primarily to the specific removal from circulation of a class of T cells with high GSH;^{3,6} and (4) GSH levels in B cells and monocytes, while not significantly different from HIV⁻ individuals, vary from the mean considerably more than in normals.⁴

One aspect of the inflammatory stress in AIDS, noted as early as 1983, is polyclonal B cell activation.⁹⁷ Several observations confirm the activated state of B cells in HIV-infected individuals, including hypergammaglobulinemia, elevated expression of B-cell activation markers, increased frequency of B lymphomas, and elevated levels of plasma IL-6.^{5,85,97-101}

In contrast to the B cells, T cells and monocytes appear to be less responsive if not anergic in HIV-infected individuals. Defects of immune function both *in vivo* and *in vitro* were observed early in AIDS research (e.g., Fauci et al.¹⁰²). This deficiency might be brought about by excessive stimulation with cytokines¹⁰³⁻¹⁰⁵ (i.e., induced anergy). Voth et al. found that PBMC from HIV-infected individuals produce lower levels of IFN- α and IFN- γ (but increased levels of TNF) in response to stimulation.⁸² Fuchs et al. found that IFN- γ production by PBMC from HIV-infected individuals inversely correlates with the serum levels of IFN- γ ,⁸⁷ as is true for IL-2 production and serum levels of neopterin.¹⁰⁶ They predict that the diminished PBMC responsiveness will be found not only for AIDS, but also for other diseases in which immune activation is present.¹⁰⁴

In addition to cytokine-induced anergy may be a low-GSH-induced anergy. The progressive loss of T-cell function in AIDS-related complex (ARC) and AIDS patients^{102,107} and the polyclonal B-cell activation correlate with the observed decrease in intracellular GSH in T cells but not B cells. In view of the sensitivity of T-cell (and other lymphocyte) function to GSH levels,^{62,75,77} the loss of GSH may partially explain the immunodeficiency in these HIV-infected individuals. As discussed above, depletions of intracellular GSH can lead to complete inhibition of mitogen-induced T-cell proliferation.

TREATMENT OF HIV INFECTION SHOULD INCLUDE GSH REPLACEMENT THERAPY

It is clear now that there is an intricate and large set of interactions between GSH levels, the production of oxidants, inflammatory cytokines, and the progression of HIV infection. These interactions are depicted in Figure 1. From these interactions, we predict that there are several loops in the network which contribute to progressive deterioration in the health of HIV-infected individuals. First, there is the autocrine stimulation of cytokines: for example, TNF stimulates its own production.^{31,108-110} Second, the stimulated production of ROIs can lead to decreased GSH levels, which lead to increased TNF sensitivity. And third, both cytokines and ROIs directly stimulate HIV; the HIV infection leads to chronically elevated levels of cytokines *in vivo*.

Increasing GSH levels can not only decrease ROIs, but also inhibit stimulation by inflammatory cytokines. Thus, this part of the network represents an excellent point for therapeutic intervention. Therapies designed to increase GSH levels in the blood could have many positive effects: restoration of general GSH levels, alleviating the observed radiation sensitivity of HIV⁺

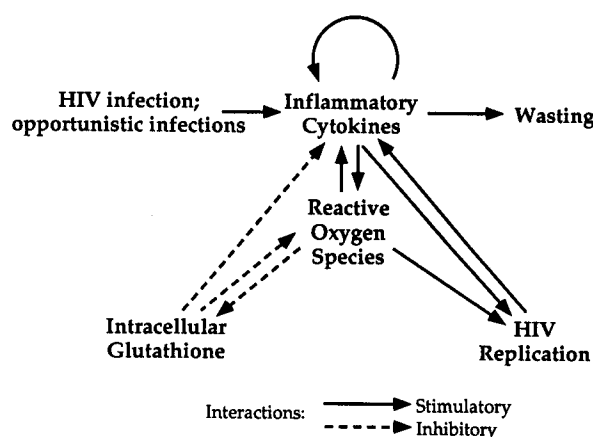


FIG. 1. Dynamics of inflammatory cytokines, ROIs, and glutathione during the progression of the HIV infection. Positive interactions (stimulations) and negative interactions (inhibitions) result in positive feedback loops between inflammatory cytokines (such as TNF), the production of ROIs, decreases in intracellular GSH, and increases in HIV replication. Opportunistic infections can pump this cycle through stimulation of the immune system; overproduction of TNF (and possibly other cytokines) may lead to progressive wasting and eventual death. See text for more detailed discussion of the various pathways.

individuals;¹¹¹ restoration of GSH levels in T cells, thereby possibly alleviating the suppressed state of these cells; inhibition of further inflammatory stimulation and production of ROIs, thereby preventing direct stimulation of virus and further production of cytokines; and reduction of TNF (cachectin¹¹²) levels, thereby potentially alleviating the distressing wasting (cachexia) that often accompanies late-stage AIDS.¹¹³

Many antioxidants are of potential use in this regard. Some compounds, which do not replenish GSH, may have beneficial effects by inhibiting ROI production and sparing extant GSH. For instance, penicillamine inhibits HIV production effectively *in vitro*,¹¹⁴ and its effectiveness as an anti-inflammatory is correlated with increasing GSH levels.¹¹⁵ It has antiviral activity in HIV-infected individuals; however, penicillamine induced transient depressions in CD4 T-cell numbers and lymphoproliferative capacity in these individuals^{116,117}. Hydrazine sulfate, an anticachectic agent used in cancer trials^{118,119} inhibits TNF *in vitro*,¹²⁰ and it inhibits both the TNF and PMA stimulation of viral transcription and viral replication (Roederer, unpublished). Auranofin, a gold-based antioxidant which quenches singlet oxygen¹²¹ and is used as an antiarthritic, specifically inhibits PMA but not TNF stimulations (Roederer, unpublished). Finally, vitamin C effectively inhibits HIV replication.^{122,123}

However, we feel that most effective compounds will be GSH prodrugs; NAC, OTC, GSH itself, and glutathione ester (GSE). NAC has a long history of use in humans, dating back 25 years. Its pharmacokinetics and safety are well-established.^{52,124-127} OTC is similar to NAC in that it is a cysteine prodrug requiring intracellular enzymatic conversion;¹²⁸ its pharmacokinetics have also been described.¹²⁹ GSH and GSE also are possible candidates: they have antiviral activity¹⁹ and can increase plasma GSH.^{130,131} As Halliwell and Cross suggest, antioxidant

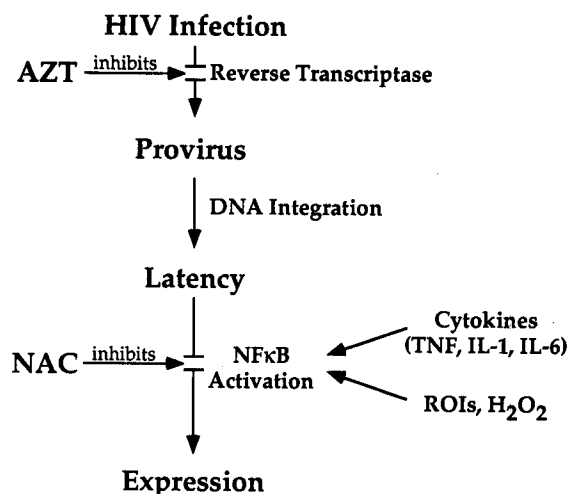


FIG. 2. NAC is a novel approach to anti-HIV therapy. NAC inhibits the host-mediated stimulation of viral transcription and production. Current therapies (AZT, DDI, DDC) are directed at inhibition of the reverse transcriptase and are ineffective in the inhibition of viral production after latency has been established.

therapy for patients is merited for drugs like vitamin C and NAC, which have extremely limited toxic effects.²¹

NAC, GSH, and GSE are effective in blocking the stimulation of acute HIV replication and the stimulation of latently infected cell lines. As of now, there are few therapies which are designed to inhibit HIV *after* it has integrated into the genome; i.e., AZT, DDI, and DDC all work by inhibiting the reverse transcriptase necessary for integration of the virus. Furthermore, owing to the mutability of HIV, resistant variants of the virus can arise during long-term treatments with these drugs. GSH replenishment therapy should be effective in extending latency by inhibiting the host-based stimulation of HIV replication (Fig. 2). Furthermore, this type of therapy may have many positive effects on the symptomatology of the HIV infection, e.g., against lymphocyte energy, immunosuppression, development of opportunistic infections, and wasting.

ACKNOWLEDGMENTS

We thank Drs. Poli, Fauci, and Baeuerle for sharing unpublished data. MR is a Senior Fellow of the Leukemia Society of America; FJTS was supported by Department of Genetics funds, and LAH was supported in part by NIH Grant CA42509.

REFERENCES

- Eck H-P, Gmunder H, Hartmann M, Petzoldt D, Daniel V, and Droge W: Low concentrations of acid-soluble thiol (cysteine) in the blood plasma of HIV-1-infected patients. *Biol Chem Hoppe-Seyler* 1989;370:101.
- Buhl R, Holroyd KJ, Mastrangeli A, Cantin AM, Jaffe HA, Wells FB, Saltini C, and Crystal RG: Systemic glutathione deficiency in symptom free HIV-seropositive individuals. *Lancet* 1989;2:1294.

- Roederer M, Staal FJT, Osada H, Herzenberg LA, and Herzenberg LA: CD4 and CD8 T cells with high intracellular glutathione levels are selectively lost as the HIV infection progresses. *Intl Immunol* 1991;3:933.
- Roederer M, Staal FJT, Raju PA, Herzenberg LA, and Herzenberg LA: The interrelationship of TNF, glutathione and AIDS. In: *Proceedings from the 3rd TNF and Related Cytokines Conference*. S. Karger-AG, Basel, 1991.
- Staal F, Roederer M, Herzenberg L, and Herzenberg L: Glutathione and immunophenotypes of T and B lymphocytes in HIV infected individuals. In: *CD5 B Cells in Development and Disease*. New York Academy of Sciences, New York, 1991.
- Staal FJT, Roederer M, Israelski DM, Bubp J, Mole LA, McShane D, Deresinski SC, Ross W, Sussman H, Raju PA, Anderson MT, Moore W, Ela SW, Herzenberg LA, and Herzenberg LA: Intracellular glutathione levels in T cell subsets decrease in HIV infected individuals. *AIDS Res Human Retroviruses* 1992; 2:311.
- Smith CV, Hansen TN, Hanson IC, and Shearer WT: Glutathione concentrations in plasma and blood are markedly decreased in HIV-infected children. *Sixth Int Conf AIDS* 1990;II:368 (abstr).
- Folks TM, Clouse KA, Justement J, Rabson A, Duh E, Kehrl JH, and Fauci AS: Tumor necrosis factor alpha induces expression of human immunodeficiency virus in a chronically infected T-cell clone. *Proc Natl Acad Sci* 1989;86:2365.
- Poli G, Bressler P, Kinter A, Duh E, Timmer WC, Rabson A, Justement JS, Stanley S, and Fauci AS: Interleukin 6 induces human immunodeficiency virus expression in infected monocytic cells alone and in synergy with tumor necrosis factor α by transcriptional and post-transcriptional mechanisms. *J Exp Med* 1990;172:151.
- Stanley SK, Bressler PB, Poli G, and Fauci AS: Heat shock induction of HIV production from chronically infected promonocytic and T cell lines. *J Immunol* 1990;145:1120.
- Lahdevirta J, Maury CPJ, Teppo A-M, and Repo H: Elevated levels of circulating cachectin/tumor necrosis factor in patients with acquired immunodeficiency syndrome. *Am J Med* 1988;85:289.
- Reddy MM, Sorrell SJ, Lange M, and Grieco MH: Tumor necrosis factor and HIV P24 antigen levels in serum of HIV-infected populations. *J AIDS* 1988;1:436.
- Mintz M, Rapaport R, Oleske JM, Connor EM, Koenigsberger MR, Denny T, and Epstein LG: Elevated serum levels of tumor necrosis factor are associated with progressive encephalopathy in children with acquired immunodeficiency syndrome. *Am J Dis Child* 1989;143:771.
- Kobayashi S, Hamamoto Y, Kobayashi N, and Yamamoto N: Serum level of TNF α in HIV-infected individuals. *AIDS* 1990;4:169.
- von Sydow M, Sonnerborg A, Gaines H, and Strannegard O: Interferon-alpha and tumor necrosis factor-alpha in serum of patients in various stages of HIV-1 infection. *AIDS Res Human Retroviruses* 1991;7:375.
- Scott-Algara D, Vuillier F, Marasescu M, de Saint Martin J, and Dighiero G: Serum levels of IL-2, IL-1 α , TNF- α , and soluble receptor of IL-2 in HIV-1-infected patients. *AIDS Res Human Retroviruses* 1991;7:381.
- Gallo P, Frei K, Rordorf C, Lazdins J, Tavolato B, and Fontana A: Human immunodeficiency virus type 1 (HIV-1) infection of the central nervous system: an evaluation of cytokines in cerebrospinal fluid. *J Neuroimmunol* 1989;23:109.
- Roederer M, Staal FJT, Raju PA, Ela SW, Herzenberg LA, and Herzenberg LA: Cytokine-stimulated HIV replication is inhibited by N-acetylcysteine. *Proc Natl Acad Sci (USA)* 1990;87:4884.
- Kalebic T, Kinter A, Poli G, Anderson ME, Meister A, and Fauci AS: Suppression of HIV expression in chronically infected mono-

- cytic cells by glutathione, glutathione ester, and N-acetylcysteine. *Proc Natl Acad Sci (USA)* 1991;88:986.
20. Mihm S, Ennen J, Pessara U, Kurth R, and Droge W: Inhibition of HIV-1 replication and NF-kappaB activity by cysteine and cysteine derivatives. *AIDS* 1991;5:497.
 21. Halliwell B and Cross CE: Commentary: reactive oxygen species, antioxidants, and acquired immunodeficiency syndrome—sense or speculation. *Arch Intern Med* 1991;151:29.
 22. Roederer M, Raju PA, Staal FJT, Herzenberg LA, and Herzenberg LA: N-acetylcysteine inhibits latent HIV expression in chronically infected cells. *AIDS Res Human Retroviruses* 1991;7:491.
 23. Staal FJT, Roederer M, Herzenberg LA, and Herzenberg LA: Intracellular thiols regulate activation of nuclear factor kappaB and transcription of human immunodeficiency virus. *Proc Natl Acad Sci (USA)* 1990;87:9943.
 24. Nabel G and Baltimore D: An inducible transcription factor activates expression of human immunodeficiency virus in T cells. *Nature* 1987;326:711.
 25. Baeuerle PA and Baltimore D: IkappaB: a specific inhibitor of the NF-kappaB transcription factor. *Science* 1988;242:540.
 26. Baeuerle PA and Baltimore D: Activation of DNA-binding activity in an apparently cytoplasmic precursor of the NF-kappaB transcription factor. *Cell* 1988;53:211.
 27. Clouse KA, Powell D, Washington I, Poli G, Strebel K, Farrar W, Barstad P, Kovacs J, Fauci AS and Folks TM: Monokine regulation of human immunodeficiency virus-1 expression in chronically infected human T cell clone. *J Immunol* 1989;142:431.
 28. Duh EJ, Maury WJ, Folks TM, Fauci AS, and Rabson AB: Tumor necrosis factor alpha activates human immunodeficiency virus type 1 through induction of nuclear factor binding to the NF-kappaB sites in the long terminal repeat. *Proc Natl Acad Sci (USA)* 1989;86:5974.
 29. Osborn L, Kunkel S, and Nabel GJ: Tumor necrosis factor alpha and interleukin 1 stimulate the human immunodeficiency virus enhancer by activation of the nuclear factor kappaB. *Proc Natl Acad Sci* 1989;86:2336.
 30. Griffin GE, Leung K, Folks TM, Kunkel S, and Nabel GJ: Activation of HIV gene expression during monocyte differentiation by induction of NF-kappa B. *Nature* 1989;339:70.
 31. Poli G, Kinter A, Justement JS, Kehrl JH, Bressler P, Stanley S, and Fauci AS: Tumor necrosis factor alpha functions in an autocrine manner in the induction of human immunodeficiency virus expression. *Proc Natl Acad Sci (USA)* 1990;87:782.
 32. Israel N, Hazan U, Alcamí A, Munier A, Arenzana-Seisdedos F, Bachelier F, Israel A, and Virelizier J-L: Tumor necrosis factor stimulates transcription of HIV-1 in human T lymphocytes, independently and synergistically with mitogens. *J Immunol* 1989;143:3956.
 33. Klebanoff SJ, Vadas MA, Harlan JM, Sparks LH, Gamble JR, Agosti JM, and Waltersdorff AM: Stimulation of neutrophils by tumor necrosis factor. *J Immunol* 1986;136:4220.
 34. Nathan CF: Neutrophil activation on biological surfaces: massive secretion of hydrogen peroxide in response to products of macrophages and lymphocytes. *J Clin Invest* 1987;80:1550.
 35. Kapp A, Zeck-Kapp G, and Blohm D: Human tumor necrosis factor is a potent activator of the oxidative metabolism in human polymorphonuclear neutrophilic granulocytes: comparison with human lymphotoxin. *J Invest Dermatol* 1989;92:348.
 36. Cross AR and Jones OTG: Enzymic mechanisms of superoxide production. *Biochim Biophys Acta* 1991;1057:281.
 37. Maly F-E: The B lymphocyte: a newly recognized source of reactive oxygen species with immunoregulatory potential. *Free Radic Res Commun* 1990;8:143.
 38. Leca G, Benichou G, Bensussan A, Mitene F, Galanaud P, and Vazquez A: Respiratory burst in human B lymphocytes. *J Immunol* 1991;146:3542.
 39. Sekkat C, Dornand J, and Gerber M: Oxidative phenomena are implicated in human T-cell stimulation. *Immunology* 1988;63:431.
 40. Benichou G, Kanellopoulos JM, Mitene F, Galanaud P, and Leca G: T-cell chemiluminescence. A novel aspect of T-cell membrane activation studied with a Jurkat tumour cell line. *Scand J Immunol* 1989;30:265.
 41. Gorog P, Pearson JD, and Kakkar VV: Generation of reactive oxygen metabolites by phagocytosing endothelial cells. *Atherosclerosis* 1988;72:19.
 42. Yamauchi N, Kuriyama H, Watanabe N, Neda H, Maeda M, and Niitsu Y: Intracellular hydroxyl radical production induced by recombinant human tumor necrosis factor and its implication in the killing of tumor cells in vitro. *Cancer Res* 1989;49:1671.
 43. Meier B, Radeke HH, Selle S, Habermehl GG, Resch K, and Sies H: Human fibroblasts release low amounts of reactive oxygen species in response to the potent phagocyte stimulants, serum-treated zymosan, N-Formyl-methionyl-leucyl-phenylalanine, Leukotriene B₄ or 12-O-tetradecanoylphorbol 13-acetate. *Biol Chem Hoppe-Seyler* 1990;371:1021.
 44. DeChatelet LR, Shirley PS, and Johnston RB: Effect of phorbol myristate acetate on the oxidative metabolism of human polymorphonuclear leukocytes. *Blood* 1976;47:545.
 45. Yoshie O, Majima T, and Saito H: Membrane oxidative metabolism of human eosinophilic cell line EoL-1 in response to phorbol diester and formyl peptide: synergistic augmentation by interferon-gamma and tumor necrosis factor. *J Leuk Biol* 1989;45:10.
 46. Dolphin D, Avramovic O, and Poulson R: Glutathione: Chemical, Biochemical, and Medical Aspects. In: *Coenzymes and Cofactors*, Vol 3, John Wiley & Sons, NY, 1989.
 47. Tanaguchi M, Hirayama K, Yamaguchi K, Tateishi N, and Suzuki M: Nutritional aspects of glutathione metabolism and function. In: *Glutathione: Chemical, Biochemical, and Medical Aspects*. D. Dolphin, O. Avramovic, and R. Poulson (eds.). John Wiley and Sons, New York, 1989, p. 645.
 48. Perchellet EM, Maatta EA, Abney NL, and Perchellet JP: Effects of diverse intracellular thiol delivery agents on glutathione peroxidase activity, the ratio of reduced/oxidized glutathione, and ornithine decarboxylase induction in isolated mouse epidermal cells treated with 12-O-tetradecanoylphorbol-13-acetate. *J Cell Physiol* 1987;131:64.
 49. Bilzer M and Lauterburg BH: Glutathione metabolism in activated human neutrophils: stimulation of glutathione synthesis and consumption of glutathione by reactive oxygen species. *Eur J Clin Invest* 1991;21:316.
 50. Zimmerman RJ, Marafino Jr. BJ, Chan A, Landre P, and Winkelhake JL: The role of oxidant injury in tumor cell sensitivity to recombinant human tumor necrosis factor in vivo. Implications for mechanism of action. *J Immunol* 1989;142:1405.
 51. Aruoma OI, Halliwell B, Hoey BM, and Butler J: The antioxidant action of N-acetylcysteine: its reaction with hydrogen peroxide, hydroxyl radical, superoxide, and hypochlorous acid. *Free Radic Biol Med* 1989;6(6):593.
 52. Burgunder JM, Varriale A, and Lauterburg BH: Effect of N-acetylcysteine on plasma cysteine and glutathione following paracetamol administration. *Eur J Clin Pharmacol* 1989;36:127.
 53. Zimmerman RJ, Chan A, and Leadon SA: Oxidative damage in murine tumor cells treated in vitro by recombinant human tumor necrosis factor. *Cancer Res* 1989;49:1644.
 54. Prescott LF, Illingsworth RN, Critchley JA, Stewart MJ, Adam RD, and Proudfoot AT: Intravenous N-acetylcysteine: the

- treatment of choice for paracetamol poisoning. *Br Med J* 1979;2:1097.
55. Prescott LF and Critchley JAJH: The treatment of acetaminophen poisoning. *Ann Rev Pharmacol Toxicol* 1983;23:87.
 56. Smilkstein MJ, Knapp GL, Kulig KW, and Rumack BH: Efficacy of oral *N*-acetylcysteine in the treatment of acetaminophen overdose. Analysis of the National Multicenter Study (1976 to 1985). *N Engl J Med* 1988;319:1557.
 57. Richman P and Meister A: Regulation of γ -glutamyl-cysteine synthetase by nonallosteric feedback inhibition by glutathione. *J Biol Chem* 1975;250:1422.
 58. Meister A: Selective modification of glutathione metabolism. *Science* 1983;220:472.
 59. Meister A: Methods for the selective modification of glutathione metabolism and study of glutathione transport. *Methods Enzymol* 1985;113:571.
 60. Yamauchi N, Watanabe N, Kuriyama H, Neda H, Maeda M, Himeno T, Tsuji Y, and Niitsu Y: Suppressing effects of intracellular glutathione on hydroxyl radical production induced by tumor necrosis factor. *Int J Cancer* 1990;46:884.
 61. Fischman CH, Udey MC, Kurtz M, and Wedner JH: Inhibition of lectin-induced lymphocyte activation by 2-cyclohexene-1-one: decreased intracellular glutathione inhibits an early event in the activation sequence. *J Immunol* 1981;127:2257.
 62. Fidelus RK and Tsan M-F: Enhancement of intracellular glutathione promotes lymphocyte activation by mitogen. *Cell Immunol* 1986;97:155.
 63. Fidelus RK, Ginouves P, Lawrence D, and Tsan M-F: Modulation of intracellular glutathione concentrations alters lymphocyte activity and proliferation. *Exp Cell Res* 1987;170:269.
 64. Fidelus RK: The generation of oxygen radicals: a positive signal for lymphocyte activation. *Cell Immunol* 1988;113:175.
 65. Suthanthiran M, Anderson ME, Sharma VK, and Meister A: Glutathione regulates activation-dependent DNA synthesis in highly purified normal human T lymphocytes stimulated via the CD2 and CD3 antigens. *Proc Natl Acad Sci (USA)* 1990; 87:3343.
 66. Messina JP and Lawrence DA: Cell cycle progression of glutathione-depleted human peripheral blood mononuclear cells is inhibited at S phase. *J Immunol* 1989;143:1974.
 67. Ballard DW, Bohnlein E, Lowenthal JW, Wano Y, Franza R, and Greene WC: HTLV-1 Tax induces cellular proteins that activate the kappaB element in the IL-2 receptor alpha gene. *Science* 1988;241:1652.
 68. Lowenthal JW, Ballard DW, Bohnlein E, and Greene WC: Tumor necrosis factor alpha induces proteins that bind specifically to kappaB-like enhancer elements and regulate interleukin 2 receptor alpha-chain gene expression in primary human T lymphocytes. *Proc Natl Acad Sci (USA)* 1989;86:2331.
 69. Shibuya H, Yoneyama M, and Taniguchi T: Involvement of a common transcription factor in the regulated expression of IL-2 and IL-2 receptor genes. *Intl Immunol* 1989;1:43.
 70. Gmunder H, Roth S, Eck HP, Gallas H, Mihm S, and Droge W: Interleukin-2 mRNA expression, lymphokine production and DNA synthesis in glutathione-depleted T cells. *Cell Immunol* 1990;130:520.
 71. Gmunder H, Eck H-P, Benninghoff B, Roth S, and Droge W: Macrophages regulate intracellular glutathione levels of lymphocytes. Evidence for an immunoregulatory role of cysteine. *Cell Immunol* 1990;129:32.
 72. Wong GHW, Elwell JH, Oberley LW, and Goeddel DV: Manganous superoxide dismutase is essential for cellular resistance to tumor necrosis factor. *Cell* 1989;58:923.
 73. Wong GHW, and Goeddel DV: Induction of manganous superoxide dismutase by tumor necrosis factor: possible protective mechanism. *Science* 1988;242:941.
 74. Wong GHW, Mchugh T, Weber R, and Goeddel DV: Tumor necrosis factor-alpha selectively sensitizes human immunodeficiency virus-infected cells to heat and radiation. *Proc Natl Acad Sci (USA)* 1991;88:4372.
 75. Hamilos DL and Wedner HJ: The role of glutathione in lymphocyte activation. I. Comparison of inhibitory effects of buthionine sulfoximine and 2-cyclohexene-1-one by nuclear size transformation. *J Immunol* 1985;135:2740.
 76. Hamilos DL, Zelarney P, and Mascali JJ: Lymphocyte proliferation in glutathione-depleted lymphocytes: direct relationship between glutathione availability and the proliferative response. *Immunopharmacology* 1989;18:223.
 77. Droge W, Pottmeyer-Gerber C, Schmidt H, and Nick S: Glutathione augments the activation of cytotoxic T lymphocytes in vivo. *Immunobiology* 1986;172:151.
 78. Schreck R, Rieber P, and Baeuerle PA: Reactive oxygen intermediates as apparently widely used messengers in the activation of the NF-kappaB transcription factor and HIV-1. *EMBO J* 1991;10:2247.
 79. Legrand-Poels S, Vaira D, Pincemail J, Van de Vorst A, and Piette J: Activation of human immunodeficiency virus type I by oxidative stress. *AIDS Res Human Retroviruses* 1990;6:1389.
 80. Frei B, Yamamoto Y, Niclas D, and Ames BN: Evaluation of an isoluminol chemiluminescence assay for the detection of hydroperoxides in human blood plasma. *Anal Biochem* 1988;175:120.
 81. Fuchs D, Jager H, Popescu M, Reibnegger G, Werner ER, Dierich MP, Kaboth W, Tilz GP, and Wachter H: Immune activation markers to predict AIDS and survival in HIV-1 seropositives. *Immunol Lett* 1990;26:75.
 82. Voth R, Rossol S, Klein K, Hess G, Schutt KH, Schroder HC, Meyer zum Buschenfelde K-H and Muller WEG: Differential gene expression of IFN-alpha and tumor necrosis factor-alpha in peripheral blood mononuclear cells from patients with AIDS related complex and AIDS. *J Immunol* 1990;144:970.
 83. Israel-Biet D, Cadranet J, Beldjord K, Andrieu J-M, Jeffrey A, and Even P: Tumor necrosis factor production in HIV-seropositive subjects. *J Immunol* 1991;147:490.
 84. Agostini C, Zambello R, Trentin L, Garbisa S, Fancia di Celle L, Bulian P, Onisto M, Poletti V, Spiga L, Rause E, Foa R, and Semenzato G: Alveolar macrophages from patients with AIDS and AIDS-related complex constitutively synthesize and release tumor necrosis factor alpha. *Am Rev Respir Dis* 1991;144:195.
 85. Breen EC, Rezai Ar, Nakajima K, Beall GN, Mitsuyasu RT, Hirano T, Kishimoto T, and Martinez-Maza O: Infection with HIV is associated with elevated IL-6 levels and production. *J Immunol* 1990;144:480.
 86. Eyster ME, Goedert JJ, Poon MC, and Preble OT: Acid-labile alpha interferon. A possible preclinical marker for the acquired immunodeficiency syndrome in hemophilia. *N Engl J Med* 1983;309:583.
 87. Fuchs D, Hausen A, Reibnegger G, Werner ER, Werner-Felmayer G, Dierich MP, and Wachter H: Interferon-gamma concentrations are increased in sera from individuals infected with human immunodeficiency virus type 1. *J AIDS* 1989;2:158.
 88. Grunfeld C, Kotler DP, Hamadeh R, Tierney A, Wang J, and Pierson RN: Hypertriglyceridemia in the acquired immunodeficiency syndrome. *Am J Med* 1989;86:27.
 89. Wachter H, Fuchs D, Hause A, Reibnegger G, and Werner ER: Neopterin as marker for activation of cellular immunity: immunological basis and clinical applications. *Adv Clin Chem* 1989;27:81.
 90. Fuchs D, Hausen A, Reibnegger G, Werner ER, Dierich MP, and

- Wachter P: Neopterin as a marker for activated cell-mediated immunity: application in HIV infection. *Immunol Today* 1988;9:150.
91. Melmed RN, Taylor JM, Detels R, Bozorgmehri M, and Fahey JL: Serum neopterin changes in HIV-infected subjects: indicator of significant pathology, CD4 T cell changes, and the development of AIDS. *J AIDS* 1989;2:70.
 92. Sheppard HW, Ascher MS, McRae B, Anderson RE, Lang W, and Allain JP: The initial immune response to HIV and immune system activation determine the outcome of HIV disease. *J AIDS* 1991;4:704.
 93. Reibnegger G, Spira TJ, Fuchs D, Wernerfermayer G, Dierich MP, and Wachter H: Individual probability for onset of full-blown disease in patients infected with human immunodeficiency virus type-1. *Clin Chem* 1991;37:351.
 94. Osmond DH, Shiboski S, Bacchetti P, Winger EE, and Moss AR: Immune activation markers and AIDS prognosis. *AIDS* 1991; 5:505.
 95. Zmuda J and Friedenson B: Changes in intracellular glutathione levels in stimulated and unstimulated lymphocytes in the presence of 2-mercaptoethanol or cysteine. *J Immunol* 1983;130:362.
 96. Eck H-P, Stahl-Hennig C, Hunsmann G, and Droge W: Metabolic disorder as early consequence of simian immunodeficiency virus infection in rhesus macaques. *Lancet* 1991;1:346.
 97. Lane HC, Masur H, Edgar LC, and Fauci AS: Abnormalities of B cell activation and immunoregulation in patients with the acquired immunodeficiency syndrome. *N Engl J Med* 1983;309:453.
 98. Lane HC and Fauci AS: Immunological abnormalities in the acquired immunodeficiency syndrome. *Ann Rev Immunol* 1985; 3:477.
 99. Ziegler JL, Beckstead JA, Volberding PA, et al: Non-Hodgkin's lymphoma in 90 homosexual men: relation to generalized lymphadenopathy and the acquired immunodeficiency syndrome. *N Engl J Med* 1984;311:565.
 100. Yarchoan R, Redfield RR, and Broder S: Mechanisms of B cell activation in patients with acquired immunodeficiency syndrome and related disorders. *J Clin Invest* 1986;78:439.
 101. Martinez-Maza O, Crabb E, Mitsuyasu RT, Fahey JL, and Giorgi JV: Infection with the human immunodeficiency virus (HIV) is associated with an in vivo increase in B lymphocyte activation and immaturity. *J Immunol* 1987;138:3720.
 102. Fauci AS, Macher AM, Longo DL, Lane HC, Rook AH, Masur H, and Gelmann EP: Acquired immunodeficiency syndrome: epidemiologic, clinical, immunologic, and therapeutic considerations. *Ann Intern Med* 1984;100:92.
 103. Stewart II WE: *The Interferon System*. Springer Verlag, New York, 1979.
 104. Fuchs D, Malkowsky M, Reibnegger G, Werner ER, Forni G, and Wachter H: Endogenous release of interferon-gamma and diminished response of peripheral blood mononuclear cells to antigenic stimulation. *Immunol Lett* 1989;23:103.
 105. Ascher MA, and Sheppard HW: AIDS as immune system activation II: the panergic imnesia hypothesis. *J AIDS* 1990;3:177.
 106. Fuchs D, Shearer GM, Boswell RN, Clerici M, Reibnegger G, Werner ER, Zajac RA, and Wachter H: Increased serum neopterin in patients with HIV-1 infection is correlated with reduced in vitro interleukin-2 production. *Clin Exp Immunol* 1990;80:44.
 107. Mitsuyasu RT: Medical aspects of HIV spectrum disease. *Psychiatr Med* 1989;7:5.
 108. Philip R, and Epstein LB: Tumour necrosis factor as immunomodulator and mediator of monocyte cytotoxicity induced by itself, gamma-interferon and interleukin-1. *Nature* 1986;323:86.
 109. Hensel G, Mannel DN, Pfizenmaier K, and Kronke M: Autocrine stimulation of TNF-alpha mRNA expression in HL-60 cells. *Lymphokine Res* 1987;6:119.
 110. Cordingley FT, Hoffbrand AV, Heslop HE, Turner M, Bianchi A, Reittie JE, Vyakarnam A, Meager A, and Brenner MK: Tumour necrosis factor as an autocrine tumour growth factor for chronic B-cell malignancies. *Lancet* 1988;1:969.
 111. Vallis KA: Glutathione deficiency and radiosensitivity in AIDS patients. *Lancet* 1991;1:918.
 112. Beutler B, Greenwald D, Hulmes JD, Chang M, Pan Y-CE, Mathison J, Ulevitch R, and Cerami A: Identity of tumour necrosis factor and the macrophage-secreted factor cachectin. *Nature* 1985;316:552.
 113. Chlebowski RT: Significance of altered nutritional status in acquired immune deficiency syndrome (AIDS). *Nutr Cancer* 1985;7:85.
 114. Chandra P and Sarin PS: Selective inhibition of replication of the AIDS-associated virus HTLV-III/LAV by synthetic D-penicillamine. *Arzneimittelforschung* 1986;36:184.
 115. Munthe E, Kass E, and Jellum E: D-Penicillamine-induced increase in intracellular glutathione correlating to clinical response in rheumatoid arthritis. *J Rheumatol* 1981;8(Suppl. 7):14.
 116. Scheib RG, Parenti DM, Simon GL, Courtless JW, Schulof RS, Sarin PS, and Chandra P: Prolonged antiviral activity of D-penicillamine in human immunodeficiency virus-infected homosexual men [letter]. *Am J Med* 1987;83:608.
 117. Schulof RS, Scheib RG, Parenti DM, Simon GL, DiGioia RA, Paxton HM, Szein MB, Chandra P, Courtless JW, Taguchi YT, Sun DK, Goldstein AL, and Sarin PS: Treatment of HTLV-III/LAV-infected patients with D-penicillamine. *Arzneimittelforschung* 1986;36:1531.
 118. Gold J: Use of hydrazine sulfate in terminal and preterminal cancer patients: results of investigational new drug (IND) study in 84 evaluable patients. *Oncology* 1975;32:1.
 119. Gold J: Hydrazine sulfate: a current perspective. *Nutr Cancer* 1987;9:49.
 120. Hughes TK, Cadet P, and Larned CS: Modulation of tumor necrosis factor activities by a potential anticachexia compound, hydrazine sulfate. *Int J Immunopharmacol* 1989;11:501.
 121. Corey EJ, Mehrotra MM, and Khan AU: Antiarthritic gold compounds effectively quench electronically excited singlet oxygen. *Science* 1987;236:68.
 122. Harakeh S, Jariwalla RJ, and Pauling L: Suppression of HIV replication by ascorbate in chronically and acutely infected cells. *Proc Natl Acad Med (USA)* 1990;87:7245.
 123. Harakeh S and Jariwalla R.J.: Comparative study of the anti-HIV activities of ascorbate and thiol-containing reducing agents in chronically infected cells. *Am J Clin Nutr* 1991;54:1231.
 124. Moldeus P, Cotgreave IA, and Berggren M: Lung protection by a thiol-containing antioxidant: N-acetylcysteine. *Respiration* 1986; 50(Suppl 1):31.
 125. Olsson B, Johansson M, Gabrielsson J, and Bolme P: Pharmacokinetics and bioavailability of reduced and oxidized N-acetylcysteine. *Eur J Clin Pharmacol* 1988;34:77.
 126. Ventresca GP, Cicchetti V, and Ferrari V: Acetylcysteine. In: *Drugs in Bronchial Mucology*. P.C. Braga, and L. Allegra (eds.), Raven Press, New York, 1989.
 127. Holdiness MR: Clinical pharmacokinetics of N-acetylcysteine. *Clin Pharmacokinet* 1991;20:123.
 128. Williamson JM and Meister A: Stimulation of hepatic glutathione formation by administration of L-2-oxothiazolidine-4-carboxylate, a 5-oxo-proline substrate. *Proc Natl Acad Sci (USA)* 1981;78:936.
 129. Porta P, Aebi S, Summer K, and Lauterburg BH: L-2-Oxothiazolidine-4-carboxylic acid, a cysteine prodrug: pharmacokinetics

- and effects on thiols in plasma and lymphocytes in human. *J Pharmacol Exp Ther* 1991;257:331.
130. Hagen TM and Jones DP: Role of glutathione transport in extrahepatic detoxication. In: *Glutathione Centennial: Molecular Properties and Clinical Applications*. N. Taniguchi, T. Higashi, Y. Sakamoto, and A. Meister (eds.), Academic Press, San Diego, 1989;423.
131. Aebi S, Assereto R, and Lauterburg BH: High-dose intravenous glutathione in man—pharmacokinetics and effects on cyst(e)ine in plasma and urine. *Eur J Clin Invest* 1991;21:103.

Address reprint requests to:
Leonard A. Herzenberg
Department of Genetics
Stanford University
Stanford, CA 94305